

## КЛИНИЧЕСКАЯ МЕДИЦИНА MEDICINE



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# Effect of Cigarette Smoke on Female Reproductive System: A Sytematic Review

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### Abstract

**Background:** The closest toxic exposure is cigarette smoke, and nowadays smokers are not only men but also women. Although the number of smokers has dropped, cigarettes remain to be an element in the development of many illnesses. Cigarette toxic substances can disrupt cellular balance, including in the reproductive system. **The aim of the study:** To find out more about the effect of smoking on female reproduction. **Materials and methods:** Articles were searched in Google Scholar, Sciencedirect, Frontiers, Pubmed and Cochrane databases between 2014-2024 with the keywords "cigarette smoke, e-cigarette, nicotine, female reproductive system, uterus, endometrium, oviduct, ovary, estrogen, folliculogenesis, angiogenesis, and GnRH." Eighteen articles met the inclusion criteria. **Results:** This research is a literature review with articles selected through inclusion criteria. The results of the data analysis showed an increase in MDA, apoptosis, VEGF, iNOS, and COX-2, as well as a decrease in the number of ovarian follicles, CYP19, YAP, GnRH, AMH, FSH, LH, estradiol, SOD, GPx, CAT, thinning in oviduct thickness and oviduct mucosal folds. **Conclusion:** Smoke of cigarettes has a variety of harmful effects, including ovotoxicants. Smoking tobacco increases oxidative stress, causes inflammation, increases apoptosis leading to follicle loss, and decreases the synthesis of estrogen, GnRH, FSH, LH, progesterone and estradiol. All this affects female reproduction. There is evidence that smoking disrupts the regulation of reproductive hormones, which affects decreased reproductive functions of the ovaries, uterus, and ovaries. Although it does not affect in vitro fertilization, smoking should still be avoided due to its harmful effects on reproductive cells and hormonal balance. Therefore, given the harmful effects associated with reproduction, it is expected that smoking patterns will decrease.

**Keywords:** cigarette smoke; female; reproduction

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**Introduction.** Smoking is known to be one of the factors of premature death that can still be prevented in the world [1]. The prevalence of smoking in adolescents from 2008 (27.7%) to 2021 (9.8%) is known to have decreased. More than 75% of men who smoke every day live in countries with a medium or high Human Development Index (HDI), while more than 53% of women who smoke regularly every day live in countries with a very high HDI. Though historically low, the majority of low HDI countries have a relatively low smoking prevalence – tens of millions of people still smoke. In extremely high HDI nations, the prevalence of female smoking peaked a few decades after that of male smoking, but in the majority of low-, medium-, and high HDI countries, it has either remained relatively low or has only moderately increased thus far [2]. The survey from 2011-2022 also showed the fastest decline in cigarette users at the age of under 40 (19.2% to 4.9%), especially in those with high incomes [3]. Although the global prevalence of smoking is declining, smoking remains a major health problem in the world [4].

The types of cigarettes currently circulating are conventional cigarettes and electronic cigarettes. Conventional cigarettes are known to have high toxicity [5]. CS induces proinflammatory cytokines interleukin-8 (IL-8), interferon gamma (INF- $\gamma$ ), interleukin  $\beta$  (IL-1 $\beta$ ), tumor necrosis factor (TNF- $\alpha$ ), inter-leukin 2 (IL-2) interleukin 6 (IL-6) leading to disease progression [6]. E-cigarettes are often considered a safe substitute for conventional cigarettes [4]. E-cigarettes contain nicotine and flavor variations that can increase toxicity significantly [7]. The heat generated initiates oxidation and breaks down the components in it and ultimately forms harmful elements when inhaled [8]. Even so,

both have the same carcinogenic and toxic risks [9].

Cigarette substances including cadmium (Cd), tar, nicotine, benzo a pyrene (BaP) induce an increase in ROS and cause oxidative stress [10]. Characterized by an increase in malondialdehyde (MDA) and a decrease in catalase (CAT), superoxide dismutase (SOD), and glutathione peroxidase (GPx). The process through which ROS accumulate in cellular macromolecules results in biomolecular damage, including lipid peroxidation in cell membranes, ATP depletion, and damage to deoxyribonucleic acid (DNA), which is quantifiable by amounts of MDA. Meiotic DNA and amino acids are reacted with by chemically active MDA during the production of proteins. Because both aldehyde groups are reactive with nucleophiles, MDA can generate adducts that harm lipids. Additionally, MDA and ROS, one of the mediators of cell functional abnormalities, have a favorable relationship [11].

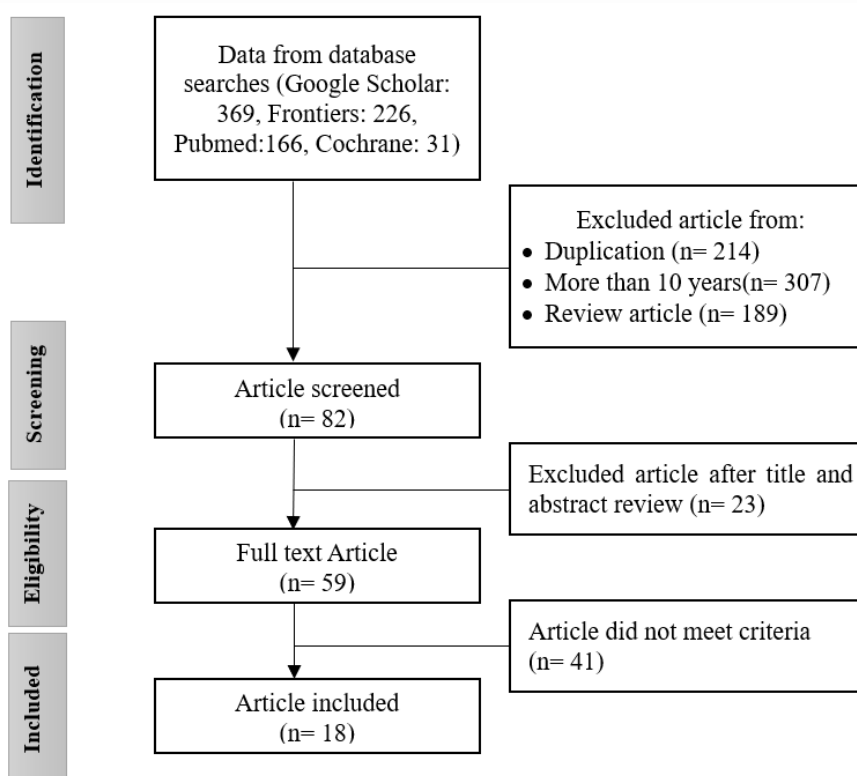
Smoking causes modifications to the cell membranes of lipid sections, induces inflammation, and vasomotor dysfunction that can lead to impaired cellular function [12]. Cigarette smoke directly damages mitochondrial respiration resulting in impaired ATP production [13]. As a result, the distribution of ATP is disrupted and the cell stops working and then undergoes death [14]. Cell death disrupts the body's homeotasis and inhibits the metabolic processes that take place [15]. In addition to active smokers, passive smokers who are exposed to cigarette smoke also have the opportunity to experience the same pain [16]. The impact of smoking in health can increase the risk of cardiovascular disease, atherosclerosis, impaired kidney function, pulmonary emphysema, and hormonal disorders [17]. In addition, cigarettes are also known to have an impact on

reproductive health by resulting in disorders of the oviduct, uterus, and ovaries [8].

**The aim of the study.** This article aims to further review the impact of cigarette smoke on the reproductive organs, especially the ovaries, oviduct, and uterus.

**Materials and Methods.** Research articles obtained from Google Scholar, Cochrane, Frontiers, and Pubmed databases with the keywords "cigarette smoke, e-

cigarette, nicotine, female reproductive, uterus, endometrium, oviduct, ovary, estrogen, folliculogenesis, angiogenesis, and GnRH". Articles reviewed are free-access articles only. The inclusion criteria are articles in 2014-2024, true experimental, quantitative research, and complete. The exclusion criteria are literature review, narrative review, and systematic review (Fig. 1).



Note: the reviewed article has gone through a search process, screening by removing the exclusion of inappropriate articles, so that articles that meet the inclusion criteria are obtained.

Fig. 1. Process of Selecting Article with PRISMA flowchart

The search yielded a total of 792 articles, of which 710 were omitted due to known duplicates, review articles and articles older than 10 years. There were selected full-text articles related to the effects of smoking on female reproduction. The articles obtained are in accordance with the inclusion criteria.

**Results.** We studied 18 articles from 2014-2024 that discussed the effects of smoking on female reproduction (Table 1). All were laboratory experimental studies with rat subjects (44.4%), mice (11.1%), and human tissue (44.4%). Materials to assess the effects of cigarettes were e-liquid (5.5%), cigarette

smoke extract (11.1%), nicotine injection (16.7%), and cigarette exposure (66.7%). All types of cigarette exposure are ovotoxicant, that is, they interfere with the normal development of follicles by decreasing the number of normal follicles, granulosa proliferation index, CYP19 and YAP expression, and serum AMH, and increasing the follicular apoptosis index especially in theca, HSCORE, iNOS expression, and COX-2. Exposure to cigarettes resulted in a decrease in antioxidants (SOD, CAT, and GPx) and an increase in markers of oxidative stress (MDA). In the fallopian tubes, smoking leads to

thinning of the mucosa and degeneration of epithelial cells. In the endometrium, smoking causes a decrease in proliferation and increase

in cell death. In addition, it also causes a decrease in the secretion of GnRH, estrogen, FSH, LH, progesterone, and estradiol.

*Beginning of Table 1*

### Review Article

Name	Ref	Method	Result	Conclusion
Chen et al, 2022	[18]	Comparing control groups and treatment with 0.005 mg and 0.5mg e-liquids.	follicles, expression of CYP19 and YAP, and estrogen was significantly reduced after exposure to e-liquid.	E-liquid affected the development of mouse reproductive.
Amalia et al, 2024	[19]	<i>Rattus norvegicus</i> divided into negative group and control group (cigarette smoke) for 28 days.	The comparison between control and negative groups resulted in decreased GnRH levels, estrogen, GDF-9, and increased MDA and apoptosis in granulosa and oocytes.	Exposure to secondhand smoke causes changes in the ovarian environment.
Kida et al, 2020	[20]	The endometrial stromal cells of 27 women were cultured with CSE-induced.	There was a decrease in endometrial proliferation and increase in cell death after CSE-induced above 1%.	Exposure to cigarette smoke affects endometrial maturation.
Chen et al, 2023	[21]	378 mouse ovaries were put into 7 groups: control, nicotine 0.05 mg/mL, <i>flavouring</i> 0.25µL/mL, PG 2.5µL/mL, VG 2.0µL/mL, LN ( <i>Low Nicotine</i> ), and HN ( <i>High Nicotine</i> ).	Nicotine has the lowest effect on the ovaries, while <i>flavouring</i> , PG, and VG cause morphological damage, oxidative balance disorders, and increased apoptosis.	There is a potential risk from the use of e-liquid to ovarian damage.
Budani et al, 2022	[22]	34 women were undergoing IVF treatment, 18 of whom were smokers and 16 were non-smokers.	CS exposure resulted in increased iNOS and COX-2 expression, as well as iNOS activity and PGE2 levels. Smoking behavior has negative correlation with serum AMH.	Cigarette smoke is ovotoxicant by increasing COX-2 as a proinflammatory that is likely to have a negative effect on the ovaries.
Rauf et al, 2022	[23]	30 rats were divided into 3 groups, group A was control, B was injected with 0.1 mg/kg nicotine for 28 days, and C was injected with nicotine and given ajwa extract.	In nicotine-injected group B, the fallopian mucosa showed flattened folds along with epithelial cell degeneration. Fibrosis and accumulation of blood vessels occur in the serous.	Nicotine can cause reversible injury to the fallopian tubes and can lead to subfertility.
Kole et al, 2019	[24]	Albino wistars were divided into group I (free air) and II (chronic cigarette smoke) for 45 days	In group II, there was decrease in the number of follicles stage, and enzymatic antioxidants (SOD, GPx, and CAT).	Cigarette smoke has relationship with ovarian reserve disorders.
Faghani et al, 2022	[25]	40 rats were put into 5 groups, A: control, B: CMC, C: nicotine 0.6mg/kg intraperitoneally, D and E: injected nicotine and given PG. Daily treatment for 30 days.	Compared to the control group, nicotine significantly lowered levels of FSH, LH, progesterone, estradiol, healthy follicles, CAT, SOD, granulosa proliferation index, theca cells, and increased tissue and serum MDA levels.	Smoking interferes with folliculogenesis by inhibiting hormone secretion, increasing markers of apoptosis, and lowering the cell proliferation index.

## Continuation of Table 1

## Review Article

Name	Ref	Method	Result	Conclusion
Susanti et al, 2020	[26]	Female <i>Mus musculus</i> were divided into 2 groups; control and the group exposed to 1 cigarette smoke for 20 days.	Smoking resulted in decrease in GnRH and increase in MDA as well as the apoptosis index of theca cells.	Cigarette smoke causes oxidative stress, a decrease in GnRH which can interfere with folliculogenesis.
Camlin et al, 2016	[27]	Female rats were exposed to cigarettes for 75 minutes every day for up to 13 days, at week 5 the mice were mated, then the research continued until the postnatal day 23.	Abnormalities were found in somatic cell proliferation and increased apoptosis, there was a decrease in the number of follicles. Increased oxidative stress was found.	Changes in the ovaries and oocytes led to subfertility seen from fewer children and longer time to conceive in cigarette-exposed mice.
Wesselink et al, 2019	[28]	Analysis of cohort studies of women and men who are planning a pregnancy in 2013-2018.	Women who are active smokers, smoke occasionally, and former smokers have been linked to decreased fertility. The highest decline occurred in women who smoked > 10 cigarettes a day.	Women smokers have lower fecundity.
Cinar et al, 2014	[29]	This comparative prospective study assessed from smoking women (43) and non-smokers (171).	The oocyte quality index, embryo development rate, fertilization rate, and pregnancy rate did not differ.	Smoking has no adverse impact on IVF outcomes.
Li et al, 2018	[30]	1000 women with PCOS, 500 women with SHS, 229 exposed and 271 unexposed were analyzed.	SHS women have high testosterone and metabolic. No difference in ovulation rates. Lower conception rates in the exposed group	Smoking spouses of PCOS women are advised to quit smoking and avoid cigarettes exposure.
Lyngsø et al, 2020	[31]	In a cohort study between 2010 and 2015, information was gathered on 1708 women and possible partners who were starting treatment cycles for frozen embryo transfer, IVF/ICSI, or IUI.	There was no correlation between CS and pregnancy or live birth in women undergoing IVF/ICSI treatment. The adjusted relative risk for smoking was 1.22 when compared to nonsmokers.	Smoking during pregnancy and reproduction therapies doesn't significantly impact clinical pregnancy or live birth, but should be discouraged.
Oladipupo et al, 2022	[32]	Smoking status was assessed using a questionnaire, and it was verified using cotinine. When ovarian reserve was assessed, AMH values below 1 ng/mL were indicative of DOR. We used NAT2 polymorphisms to determine the acetylator status. Both age and PCOS were considered. NAT2 and smoking's impact on ovarian reserve were evaluated by regression analysis.	comparing current smokers to never smokers, the infertility increased by 41.8%.	There is correlation between current smokers and an elevated risk of infertility.



End of Table 1

## Review Article

Name	Ref	Method	Result	Conclusion
Kim et al, 2018	[33]	Female rats were exposed to cigarettes 28 hours a week for 4 weeks.	In the uterus, there is an increase in CXCR4, MMP9, and $E\alpha$ . In the ovaries, the same marker increase also occurs.	CS induces uterine and ovarian abnormalities in diabetic mice.
Souza et al, 2023	[34]	Pregnant and lactating rats were exposed to 2 mg/BB cigarette smoke every day. The first offspring (F1) is mated and the second offspring (F2) is obtained.	There is a decrease in brain cell size and apoptosis in F1. In F2, there is an increase in anogenital ovaries.	Prenatal exposure to nicotine causes transgeneration in the pituitary-gonadal mice.
Konstantinidou et al, 2021	[35]	CCs from 10 donors, 5 of whom are smokers and others are not.	In smokers, an increase in IL6 and oxidative damage was found.	Imbalance triggered by cigarettes.

**Discussion****Cigarette Smoke against Ovaries**

The toxicity of cigarettes can lead to abnormalities at the cellular level, in tissues and in organs [36]. The accumulation of cigarettes in the body triggers an excessive increase in ROS and causes an imbalance called oxidative stress [24]. The mechanism of increased ROS in cellular biomolecules causes biomolecular damage that triggers the formation of malondialdehyde (MDA) [37]. As a substance that can characterize the activity of free radicals in cells, malondialdehyde (MDA) is a useful tool for identifying oxidative stress brought on by free radicals. The reactivity of both aldehyde groups to nucleophiles allows MDA to form additions that result in damage to lipids and become mediators of reproductive disorders [38].

Ovarotoxic nicotine can damage the structure of ovarian cell membranes due to low FSH stimulation of follicles, causing follicles to atrophy and eventually degenerate [39]. Low FSH levels trigger FOXO to enter the cell nucleus and induce apoptosis by releasing BCL2 and FAS ligands. Furthermore, damage occurs to mitochondria, which results in the release of cytochrome C. Cytochrome C will bind to Apaf-1, which causes the apoptosome to modulate caspase 9 into caspase 3, resulting in DNA fragmentation and apoptosis. As a result, follicles are damaged in folliculogenesis

and DNA oxidation occurs in follicles and corpus luteum [40].

Yes-associated protein (YAP) plays an important role in hippocampus signaling to trigger granulosa cell proliferation (GCs), maintenance of normal ovarian function, and follicle development, which is known to decrease when exposed to cigarettes [18]. A decrease in YAP followed by a decrease in CYP19 will inhibit estrogen synthesis [41]. In addition, cigarettes induce the production of pro-inflammatory cytokines such as  $TNF-\alpha$ , NF-kB, IL17A, and IL1B [42]. Necrosis, as one of the inflammatory responses, results in plasma membrane rupture and lysis, as well as spillage of intracellular contents into surrounding tissues causing tissue damage [43]. In this study, the tissue damage in question is follicular atresia or follicle failure to develop [44]. The results of this study correspond to Li et al (2020): cigarette exposure causes follicle loss by inhibiting autophagy and pyroptosis activation [45].

Follicular damage means mitochondrial damage to granulosa cells, consequently triggering a sustained autophagy reaction and ending in granulosa cell death resulting in a reduced number of follicles [39]. A decrease in the number of primary, primordial, and de Graaf follicles affects estrogen production and inhibits ovulation [46]. This is in accordance with the results of research by Kole et al (2020),

cigarette smoke is known to cause a decrease in the number of follicles at all stages [24]. Wesselink et al 2019 found women smokers had low fecundity. Cigarettes were found to have no negative effects on IVF, as evidenced by the fertilization rate which was no different from non-smokers[29]. However, in the second offspring (F2), it was found that the rat cubs experienced a decrease in brain function [34].

The loss of ovum at all stages is a clear indication that exposure to cigarette toxicants alters the ovaries' cell structure. The decrease in ovum was brought about by the activation of pro-inflammatory cytokines and oxidative stress indicators by CS. This occurrence causes cell damage, particularly to the GCs that make up the ovaries, which impairs the ovaries' ability to secrete hormones.

#### **Cigarette Smoke against Reproductive Hormones**

Oxidative stress from cigarette smoke can result in impaired GnRH performance, resulting in failure of secretion and synthesis of FSH and LH [25]. When GnRH does not secrete FSH and LH, there is an inhibitory process in folliculogenesis that has an impact on decreasing estrogen production by suppressing 21 hydroxylases [47]. Estrogen is needed by the body for the ovulation process, where hormone levels will be normal if follicular development takes place normally [46]. Toxic substances in cigarette smoke will increase mRNA expression in VEGF which will modulate prostaglandins, as a result progesterone synthesis is inhibited and insufficiency occurs in the corpus luteum [48]. Corpus luteum insufficiency, also called luteal phase disorder, is a manifestation of ovarian failure where no follicles develop and eventually anovulation occurs. This condition can cause menstrual abnormalities [49].

Chen et al, (2022) found that there is a decrease in CYP19 which results in a decrease in estradiol synthesis and leads to hyperandrogenism and the formation of hemorrhagic cystic follicles [50]. Exposure to CS in the ovaries causes follicles to form abnormally and creates an imbalance in the hormones that leads to a decrease in the release

of estrogen. Low estrogen causes LH to fall and FSH to stay at its highest. This has an impact on folliculogenesis later on. Therefore, reproductive diseases that are mutually exclusive arise from exposure to CS.

#### **Cigarette Smoke against Oviduct**

Rauf et al (2022) found an incidence of fallopian tube mucosa flattening after rats were injected with nicotine. His other research also found that nicotine injection can result in thinning of the wall thickness of the ampulla and isthmus, as well as the height of the epithelium of oviduct [23]. Smoking can increase inflammation, as seen with IL-6, TNF- $\alpha$ , and IL-8 which are increased [51]. Inflammation results in injury to cells that require self-repair mechanisms through proliferation [52]. Estrogen functions in the immune system of the oviduct mucosa by regulating S100A8, thereby inhibiting inflammation [53]. However, low estrogen levels due to nicotine exposure will inhibit cell proliferation which leads to cell loss and thinning of the isthmus layer [54]. Thinning of the smooth muscle layer of the fallopian tubes can decrease peristaltic movements that serve to help the movement of the ovum towards the uterus [55]. It is well known that exposure to toxins raises proinflammatory cytokines. One of the harmful effects of cigarette smoking is a drop in estrogen, which also causes inflammation, which in turn causes cell loss. Microscopic examination therefore reveals a weakening of the structural layer that constitutes the oviduct. These anomalies increase the chance of an ectopic pregnancy by preventing fertilization products from reaching the endometrium.

#### **Cigarette Smoke against Uterus**

There was a decrease in endometrial proliferation and an increase in cell death after CSE-induced (Cigarette Smoke Extract) above 1% and 0.025% CSE treatment resulted in an increase in VEGF through accumulation of factor-1 $\alpha$ . CSE-induced oxidative stress plays a role in the release of VEGF [56]. Increased VEGF is an adaptive response to the presence of oxidative stress [57]. VEGF is essential for placentation and implantation, however when the amount is excessive, it can inhibit normal

angiogenesis through over-stimulation of blood vessels and causing disruption of the structure of blood vessels [58]. CSE suppresses various regeneration functions in the endometrium, such as migration, self-repair, and pluripotency through SERPINB2 activation that can affect the implantation process [59]. Experimental animals exposed to e-cigarettes are known to have almost no embryo implantation sites despite the presence of high progesterone [60]. Angiogenesis and proliferation are highly impacted by the reduced estrogen brought on by CS, and this is where disruptions will arise. Endometrial thickness will decrease as a result of an increase in cell death (CD) in the endometrium after a reduction in proliferation. Following this thinning, there is a reduction in vascularization, which lowers the capacity of the conception products to be implanted.

**Conclusion.** The toxic effects of cigarette smoke are multifaceted, including ovotoxicants. Cigarette smoke affects female reproduction by increasing oxidative stress, triggering inflammation, increasing apoptosis to follicle loss and decreasing the synthesis of estrogen, GnRH, FSH, LH, progesterone, and estradiol. Smoking is known to damage the regulation of reproductive hormones that affect the disruption of reproductive function of the ovaries, uterus, and oviduct. Although it has no effect on IVF, smoking should still be avoided given its adverse effects on reproductive cells and hormonal balance. Therefore, it is expected that there will be a decrease in cigarette consumption patterns considering the adverse effects caused by reproduction.

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### Conflict of interests

*The authors have no conflict of interest to declare.*

### Reference

1. Kim J, Song H, Lee J, et al. Smoking and passive smoking increases mortality through mediation effect of cadmium exposure in the United States. *Scientific Reports*. 2023;13(1):3878. DOI: <https://doi.org/10.1038/s41598-023-30988-z>
2. Shin H, Park S, Yon H, et al. Estimated prevalence and trends in smoking among adolescents in South Korea, 2005–2021: a nationwide serial study. *World Journal of Pediatrics*. 2023;19(4):366-377. DOI: <https://doi.org/10.1007/s12519-022-00673-8>
3. Meza R, Cao P, Jeon J, et al. Trends in US Adult Smoking Prevalence, 2011 to 2022. *JAMA Health Forum*. 2023;4(12):e234213. DOI: <https://doi.org/10.1001/jamahealthforum.2023.4213>
4. Dai X, Gakidou E, Lopez AD. Evolution of the global smoking epidemic over the past half century: strengthening the evidence base for policy action. *Tobacco Control*. 2022;31:129-137. DOI: <https://doi.org/10.1136/tobaccocontrol-2021-056535>
5. Yayan J, Franke KJ, Biancosino C, et al. Comparative systematic review on the safety of e-cigarettes and conventional cigarettes. *Food and Chemical Toxicology*. 2024;185:114507. DOI: <https://doi.org/10.1016/j.fct.2024.114507>
6. Zhong M, Zou M, Yao Y, et al. Induction and Modulation of EVs by Cigarette Smoke and Their Relevance in Lung Disease: Recent Advances. *Journal of Respiration*. 2023;3(4):164-177. DOI: <https://doi.org/10.3390/jor3040016>
7. Diaz MC, Silver NA, Bertrand A, et al. Bigger, stronger and cheaper: growth in e-cigarette market driven by disposable devices with more e-liquid, higher nicotine concentration and declining prices. *Tobacco Control*. 2025;34:65-70. DOI: <http://dx.doi.org/10.1136/tc-2023-058033>
8. Montjean D, Pagé MHG, Bélanger MC, et al. An Overview of E-Cigarette Impact on Reproductive Health. *Life*. 2023;13(3):827. DOI: <https://doi.org/10.3390/life13030827>
9. Abbott AJ, Reibel YG, Arnett MC, et al. Oral and Systemic Health Implications of Electronic Cigarette Usage as Compared to Conventional Tobacco Cigarettes: A review of the literature. *Journal of Dental Hygiene*. 2023;97(4):21-35.
10. Seo YS, Park JM, Kim JH, et al. Cigarette Smoke-Induced Reactive Oxygen Species Formation: A Concise Review. *Antioxidants*. 2023;12(9):1732. DOI: <https://doi.org/10.3390/antiox12091732>
11. Kasililika AG, Odugogbe ATA, Dairo MD, et al. Lifestyle and oxidative stress status in infertile women in Dar es Salaam, Tanzania:



comparative cross-sectional study. Middle East Fertility Society Journal. 2021;26:33. DOI: <https://doi.org/10.1186/s43043-021-00081-4>

12. Mallah MA, Soomro T, Ali M, et al. Cigarette smoking and air pollution exposure and their effects on cardiovascular diseases. Frontiers in Public Health. 2023;11:967047. DOI: <https://doi.org/10.3389/fpubh.2023.967047>

13. Decker ST, Matias AA, Bannon ST, et al. Effects of cigarette smoke on in situ mitochondrial substrate oxidation of slow- and fast-twitch skeletal muscles. Life Sciences. 2023;315:121376. DOI: <https://doi.org/10.1016/j.lfs.2023.121376>

14. Bai X, Wang Y, Luo X, et al. Cigarette tar accelerates atherosclerosis progression via RIPK3-dependent necroptosis mediated by endoplasmic reticulum stress in vascular smooth muscle cells. Cell Communication and Signaling. 2024;22:41. DOI: <https://doi.org/10.1186/s12964-024-01480-6>

15. Chen X, Cai Q, Liang R, et al. Copper homeostasis and copper-induced cell death in the pathogenesis of cardiovascular disease and therapeutic strategies. Cell Death and Disease. 2023;14:105. DOI: <https://doi.org/10.1038/s41419-023-05639-w>

16. Qin GQ, Chen L, Zheng J, et al. Effect of passive smoking exposure on risk of type 2 diabetes: a systematic review and meta-analysis of prospective cohort studies. Frontiers in Endocrinology. 2023;14:1195354. DOI: <https://doi.org/10.3389/fendo.2023.1195354>

17. Cui J, Wang Y. Premature ovarian insufficiency: a review on the role of tobacco smoke, its clinical harm, and treatment. Journal of Ovarian Research. 2024;17:8. DOI: <https://doi.org/10.1186/s13048-023-01330-y>

18. Chen T, Wu M, Dong Y, et al. Effect of e-cigarette refill liquid on follicular development and estrogen secretion in rats. Tobacco Induced Diseases. 2022;20:36. DOI: <https://doi.org/10.18332/tid/146958>

19. Amalia A, Hendarto H, Mustika A. Nigella Sativa Ameliorates Folliculogenesis Disorders Due to Exposure to Cigarette Smoke through GnRH, MDA Expression, Estrogen Expression, GDF-9 Expression, Apoptosis Expression, and Ovarian Follicles. Journal of Medicinal and Pharmaceutical Chemistry Research. 2024;6(7):997-1009. DOI: <https://doi.org/10.48309/JMPCR.2024.444235.1122>

20. Kida N, Nishigaki A, Kakita-Kobayashi M, et al. Exposure to cigarette smoke affects

endometrial maturation including angiogenesis and decidualization. Reproductive Medicine and Biology. 2021;20(1):108-118. DOI: <https://doi.org/10.1002/rmb2.12360>

21. Chen T, Wu M, Dong Y, et al. Ovarian toxicity of e-cigarette liquids: Effects of components and high and low nicotine concentration e-cigarette liquid in vitro. Tobacco Induced Diseases. 2023;21:128. DOI: <https://doi.org/10.18332/tid/170631>

22. Budani MC, Gallorini M, Elsallabi O, et al. Cigarette smoke is associated with up-regulation of inducible NOS and COX-2 protein expression and activity in granulosa cells of women undergoing in vitro fertilization. Reproductive Toxicology. 2022;113:128-135. DOI: <https://doi.org/10.1016/j.reprotox.2022.08.013>

23. Rauf F, Suhail M, Sohail A, et al. Histological Effects of Ajwa on Oviduct after Nicotine Induced Toxicity in Adult Albino Rats. Esculapio-Journal of Services Institute of Medical Sciences. 2022;18(3):381-385. DOI: <https://doi.org/10.51273/esc22.2518328>

24. Kole E, Ozkan S, Eraldemir C, et al. Effects of melatonin on ovarian reserve in cigarette smoking: an experimental study. Archives of Medical Science. 2020;16(6):1376-1386. DOI: <https://doi.org/10.5114/aoms.2019.89409>

25. Faghani M, Saedi S, Khanaki K, et al. Ginseng alleviates folliculogenesis disorders via induction of cell proliferation and downregulation of apoptotic markers in nicotine-treated mice. Journal of Ovarian Research. 2022;15:14. DOI: <https://doi.org/10.1186/s13048-022-00945-x>

26. Susanti E, Sudiana IK, Hendarto H. Smoke effects of disturbances folliculogenesis (mda, gnRh, Hsp70, apoptosis, and follicles) in ovarian on mice balb / C. Journal of International Dental and Medical Research. 2020;13(2):774-777.

27. Camlin NJ, Sobinoff AP, Sutherland JM, et al. Maternal Smoke Exposure Impairs the Long-Term Fertility of Female Offspring in a Murine Model1. Biology of Reproduction. 2016;94(2):1-12. DOI: <https://doi.org/10.1095/biolreprod.115.135848>

28. Wesselink AK, Hatch EE, Rothman KJ, et al. Prospective study of cigarette smoking and fecundability. Human Reproduction. 2019;34(3):558-567. DOI: <https://doi.org/10.1093/humrep/dey372>

29. Cinar O, Dilbaz S, Terzioglu F, et al. Does cigarette smoking really have detrimental effects on outcomes of IVF? European Journal of

Obstetrics and Gynecology and Reproductive Biology. 2014;174:106-110. DOI: <https://doi.org/10.1016/j.ejogrb.2013.12.026>

30. Li J, Wu Q, Wu XK, et al. Effect of exposure to second-hand smoke from husbands on biochemical hyperandrogenism, metabolic syndrome and conception rates in women with polycystic ovary syndrome undergoing ovulation induction. *Human Reproduction*. 2018;33(4):617-625. DOI: <https://doi.org/10.1093/humrep/dey027>,

31. Lyngsø J, Kesmodel US, Bay B, et al. Female cigarette smoking and successful fertility treatment: A Danish cohort study. *Acta Obstetrica et Gynecologica Scandinavica*. 2021;100(1):58-66. DOI: <https://doi.org/10.1111/aogs.13979>

32. Oladipupo I, Ali T, Hein DW, et al. Association between cigarette smoking and ovarian reserve among women seeking fertility care. *PLoS ONE*. 2022;17(12):e0278998. DOI: <https://doi.org/10.1371/journal.pone.0278998>

33. Kim SM, Hwang KA, Go RE, et al. Exposure to cigarette smoke via respiratory system may induce abnormal alterations of reproductive organs in female diabetic rats. *Environmental Toxicology*. 2019;34(1):13-21. DOI: <https://doi.org/10.1002/tox.22652>

34. Souza GS, Freitas IMM, Souza JC, et al. Transgenerational effects of maternal exposure to nicotine on structures of pituitary-gonadal axis of rats. *Toxicology and Applied Pharmacology*. 2023;468:116525. DOI: <https://doi.org/10.1016/j.taap.2023.116525>

35. Konstantinidou F, Budani MC, Sarra A, et al. Impact of Cigarette Smoking on the Expression of Oxidative Stress-Related Genes in Cumulus Cells Retrieved from Healthy Women Undergoing IVF. *International Journal of Molecular Sciences*. 2021;22(23):13147. DOI: <https://doi.org/10.3390/ijms222313147>

36. Liang G, He Z, Chen Y, et al. Existence of multiple organ aging in animal model of emphysema induced by cigarette smoke extract. *Tobacco Induced Diseases*. 2022;20:02. DOI: <https://doi.org/10.18332/tid/143853>

37. Mas-Bargues C, Escriva C, Dromant M, et al. Lipid peroxidation as measured by chromatographic determination of malondialdehyde. Human plasma reference values in health and disease. *Archives of Biochemistry and Biophysics*. 2021;709:108941. DOI: <https://doi.org/10.1016/j.abb.2021.108941>

38. Kurniati ID, Nugraheni DM. Efektivitas Pemberian Ekstrak Buah Kersen (*Muntingia calabura*) terhadap Rasio Berat Testis

pada Tikus yang Dipapar Asap Rokok. *Medica Arteriana*. 2019;1(1):15-21.

39. Talakua FC, Unitly AJA. Efek Pemberian Ekstrak Etanol Rumput Kebar (*Bhiophytum petersianum* Klotzsch) terhadap Peningkatan Jumlah Folikel pada Ovarium Tikus *Rattus Norvegicus* Terpapar Asap Rokok. *Biofaal Journal*. 2020;1(2):74-84. DOI: <https://doi.org/10.30598/biofaal.v1i2pp74-84>

40. Zayani N, Susanto BNA, Solihati S. Efek Pemberian Suspensi Buah Zuriat (*Hyphaene thebaica*) terhadap Morfometri Ovarium Mencit (*Mus musculus* L.) yang Terpapar Asap Rokok. *Jurnal Ilmiah Biosaintropis (Bioscience-Tropic)*. 2023;8(2):65-76. DOI: <https://doi.org/10.33474/e-jbst.v8i2.514>

41. Liu T, Huang Y, Lin H. Estrogen disorders: Interpreting the abnormal regulation of aromatase in granulosa cells (Review). *International Journal of Molecular Medicine*. 2021;47(5):73. DOI: <https://doi.org/10.3892/ijmm.2021.4906>

42. Kumar S, Parveen S, Swaroop S, et al. TNF- $\alpha$  and MMPs mediated mucus hypersecretion induced by cigarette smoke: An in vitro study. *Toxicology in Vitro*. 2023;92:105654. DOI: <https://doi.org/10.1016/j.tiv.2023.105654>

43. Khalid N, Azimpouran M. Necrosis. In: *Treasure Island. StatPearls*; 2023.

44. Guzel EE, Kaya N, Tektemur A, et al. Chronic effects of maternal tobacco-smoke exposure and/or  $\alpha$ -lipoic acid treatment on reproductive parameters in female rat offspring. *Systems Biology in Reproductive Medicine*. 2020;66(6):387-399. DOI: <https://doi.org/10.1080/19396368.2020.1815248>

45. Li F, Ding J, Cong Y, et al. Trichostatin A alleviated ovarian tissue damage caused by cigarette smoke exposure. *Reproductive Toxicology*. 2020;93:89-98. DOI: <https://doi.org/10.1016/j.reprotox.2020.01.006>

46. Tuttle AM, Stämpfli M, Foster WG. Cigarette Smoke Causes Follicle Loss in Mice Ovaries at Concentrations Representative of Human Exposure. *Human Reproduction*. 2019;24(6):1452-1459. DOI: <https://doi.org/10.1093/humrep/dep023>

47. Casati L, Ciceri S, Maggi R, Bottai D. Physiological and pharmacological overview of the gonadotropin releasing hormone. *Biochem Pharmacol [Internet]*. 2023 Jun;212:115553. DOI: <https://doi.org/10.1016/j.bcp.2023.115553>

48. Musanejad E, Haghpanah T, Mirzaie V, et al. Effects of Ethanol and Nicotine Co-

Administration on Follicular Atresia and Placental Histomorphology in The First-Generation Mice Pups during Intrauterine Development and Lactation Periods. *Toxicology Reports*. 2021;8:793-803. DOI: <https://doi.org/10.1016/j.toxrep.2021.03.033>

49. Pizzorno JE, Murray MT. Textbook of Natural Medicine, 5th ed. Churchill Livingstone: Elsevier; 2021. DOI: <https://doi.org/10.1016/C2015-0-02243-2>

50. Ashraf S, Rasool SUA, Nabi M, et al. Impact of rs2414096 polymorphism of CYP19 gene on susceptibility of polycystic ovary syndrome and hyperandrogenism in Kashmiri women. *Scientific Reports*. 2021;11(1):12942. DOI: <https://doi.org/10.1038/s41598-021-92265-1>

51. Benowitz NL, Helen GST, Nardone N, et al. Twenty-Four-Hour Cardiovascular Effects of Electronic Cigarettes Compared With Cigarette Smoking in Dual Users. *Journal of the American Heart Association*. 2020;9:23. DOI: <https://doi.org/10.1161/JAHA.120.017317>

52. Landén NX, Li D, Stähle M. Transition from inflammation to proliferation: a critical step during wound healing. *Cellular and Molecular Life Sciences*. 2016;73(20):3861-3885. DOI: <https://doi.org/10.1007/s00018-016-2268-0>

53. Li X, Cao G, Yang H, et al. S100A8 expression in oviduct mucosal epithelial cells is regulated by estrogen and affects mucosal immune homeostasis. *PLoS ONE*. 2021;16(11):e0260188. DOI: <https://doi.org/10.1371/journal.pone.0260188>

54. Yu K, Huang ZY, Xu XL, et al. Estrogen Receptor Function: Impact on the Human Endometrium. *Frontiers in Endocrinology*. 2022;13:827724. DOI: <https://doi.org/10.3389/fendo.2022.827724>

55. Yuan S, Wang Z, Peng H, et al. Oviductal motile cilia are essential for oocyte pickup but dispensable for sperm and embryo transport. *Proceedings of the National Academy of Sciences*. 2021;118(22):e2102940118. DOI: <https://doi.org/10.1073/pnas.2102940118>

56. Behl T, Kotwani A. Exploring the various aspects of the pathological role of vascular endothelial growth factor (VEGF) in diabetic retinopathy. *Pharmacological Research*. 2015;99:137-148. DOI: <https://doi.org/10.1016/j.phrs.2015.05.013>

57. Rossino MG, Lulli M, Amato R, et al. Oxidative Stress Induces a VEGF Autocrine Loop in the Retina: Relevance for Diabetic Retinopathy.

*Cells*. 2020;9(6):1452. DOI: <https://doi.org/10.3390/cells9061452>

58. Bansal R, Ford B, Bhaskaran S, et al. Elevated Levels of Serum Vascular Endothelial Growth Factor-A Are Not Related to NK Cell Parameters in Recurrent IVF Failure. *Journal of Reproduction and Infertility*. 2017;18(3):280-287.

59. Park SR, Kim SK, Kim SR, et al. Effects of smoking on the tissue regeneration-associated functions of human endometrial stem cells via a novel target gene SERPINB2. *Stem Cell Research and Therapy*. 2022;13(1):404. DOI: <https://doi.org/10.1186/s13287-022-03061-1>

60. Wetendorf M, Randall LT, Lemma MT, et al. E-Cigarette Exposure Delays Implantation and Causes Reduced Weight Gain in Female Offspring Exposed In Utero. *Journal of the Endocrine Society*. 2019;3(10):1907-1916. DOI: <https://doi.org/10.1210/js.2019-00216>

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